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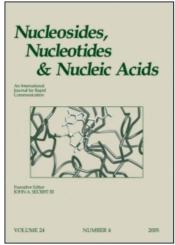
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SYNTHESIS OF 2',3',6'-TRIDEOXY-β-D-erythro-HEXOPYRANOSYL NUCLEOSIDES EMPLOYING INTRAMOLECULAR GLYCOSYLATION AS A KEY STEP

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ABSTRACT: 2',3'-Dideoxy- β -D-*erythro*-hexopyranosyl pyrimidine nucleosides were synthesized in a stereoselective manner utilizing the intramolecular pyrimidine delivery method. The nucleosides obtained from this reaction were further transformed into its 6'-deoxy derivative, which is a promising synthetic intermediate for amicetin family antibiotics.

1-(2,3,6-Trideoxy- β -D-*erythro*-hexopyranosyl)cytosine is a common skeleton of the amicetin group of antibiotics (FIG. 1), obtained from the fermentation broth of various species of *Streptomyces* and *Aethrobactor*. Amicetin was isolated in the early 1950s by several groups.¹ At present, a total of about ten compounds bearing closely related structures have been found.² As for the synthetic approach to the hexopyranosyl nucleoside moiety, Stevens *et al.*³ accomplished the first practical synthesis using the Hilbert-Johnson procedure in which a labile chloro sugar was employed as a glycosyl donor. In recent years, for other purposes, several 2',3'-deoxyhexopyranosyl nucleosides have been prepared using the Vorbrüggen coupling, which gave the desired deoxy- β -nucleosides in satisfactory yields with reasonable stereoselectivity.⁴ However, because it is often difficult to remove the contaminating undesired α -anomers, reactions with complete stereoselectivity for the β -anomer are still required. As a solution for the problem of an anomeric selectivity, we developed an intramolecular glycosylation method, in which a pyrimidine base is delivered from C-5 in pentofuranoses to the anomeric position. Using this methodology, we have demonstrated its wide applicability to the syntheses of various

This paper is dedicated to the memory of Professor Tujiaki Hata.

FIG. 1 Structure of amicetin family antibiotics

deoxynucleoside derivatives.⁵ In this paper, we describe a further extension of this method to the stereoselective synthesis of 2',3'-dideoxy-β-hexopyranosyl nucleosides.

RESULTS AND DISCUSSION

The substrates for the intramolecular glycosylation were prepared as illustrated in Scheme 1. Commercially available tri-O-acetyl-D-glucal (1) was converted into methyl 2,3-dideoxy-D-erythro-hexopyranoside (2) according to Ferrier's procedure.⁶ After conversion of 2 into thioglycoside 3 using Nicolaou's conditions,⁷ a 4-O-benzyl derivative 4a was prepared by a sequence involving deacetylation, trityl-protection of the 6-hydroxyl, benzyl-protection of the 4-hydroxyl, and removal of the trityl group under acidic conditions. Similarly, a 4-O-p-methoxybenzyl(MPM) derivative 4b was obtained by a sequence involving deacetylation, 4,6-O-p-methoxybenzylidene-protection, and reductive cleavage of the benzylidene acetal with a combination of NaBH₃CN-Me₃SiCl.⁸ The resulting thioglycosides 4a, 4b were treated with NaH and then reacted with 2-chloro-4-methoxypyrimidine,⁹ leading to the substrates of the intramolecular glycosylation, 5a and 5b.

In our previous study, we have disclosed a suitable promoter for this intramolecular glycosylation is $Me_2S(SMe)BF_4$, 5a which activates the thioglycosides $\mathbf{5a}$, $\mathbf{5b}$ to give rise to disulfide and an oxonium intermediate (SCHEME 2). Subsequently, the N-1 atom of the pyrimidine would capture the cation to form a cyclic pyrimidinium intermediate. By addition of aq. NaOH, the cyclic intermediate would be hydrolyzed to yield the desired β -nucleoside. When the 4-O-benzyl substrate $\mathbf{5a}$ was used in this intramolecular glycosylation, the β -nucleoside $\mathbf{6a}$ was obtained in 56% yield, accompanied by 23% of a C-1 hydrolyzed by-product $\mathbf{7a}$. Several reaction conditions were investigated in order to suppress the hydrolysis at C-1 (solvent, temperature, additives for stabilizing the pyrimidinium intermediate, cation effects during the hydrolysis, etc.) but none proved to be better than the original conditions. However, it was found that a great improvement lay

$$\begin{array}{c} \text{AcO} \\ \text{AcO} \\ \text{AcO} \\ \text{1} \\ \hline \\ \text{85\% (two steps)} \end{array} \begin{array}{c} \text{1. MeOH} \\ \text{BF}_3 \cdot \text{OEt}_2 \\ \text{AcO} \\ \text{AcO} \\ \text{AcO} \\ \text{AcO} \\ \text{AcO} \\ \text{AcO} \\ \text{BF}_3 \cdot \text{OEt}_2 \\ \text{81\%} \end{array} \begin{array}{c} \text{AcO} \\ \text{AcO} \\ \text{AcO} \\ \text{SPh} \\ \text{AcO} \\ \text{3} \\ \end{array}$$

- i. NaOH/MeOH
- ii. TrCl/Py; 83% (from 3)
- iii. BnBr, NaH/DMF
- iv. CF₃CO₂H/BuOH, 4a 88% (two steps)
- v. p-MeOC₆H₄CH(OMe)₂, HBF₄·OEt₂/DMF; 94% (from 3)
- vi. NaBH3CN, Me3SiCl/MeCN; 4b 87%

SCHEME 1

SCHEME 2

in the use of an MPM group as the protecting group at the 4-O position instead of the benzyl group. That is, with starting from the substrate 5b, the yield of the desired β -nucleoside 6b increased to 79%.

Finally, deoxygenation at the 6' position and removal of the 4'-O protecting groups were examined (SCHEME 3). The deoxygenation was first attempted in desulfurization of the 6'-phenylthio derivative. According to Hata's protocol, 10 6a was treated with PhSSPh-Bu₃P in pyridine, affording 6'-phenylthio derivative 8a in 98% yield. Desulfurization was then undertaken using Raney Ni (W2) to produce 6'-deoxy-4'deprotected derivative 11 in one step, though the yield was only 26%. Attempts to increase the amount of 11 failed, probably due to adsorption of Raney Ni. Therefore, an alternate route via radical reduction of a 6'-iodo derivative was examined. Treatment of 6a with I₂-PPh₃ in pyridine provided 6'-iodo derivative 9a, which was reduced with Bu₃SnH in the presence of AIBN in refluxing toluene to give rise to 6'-deoxy nucleoside 10a in a moderate overall yield (48%). Next, removal of the 4'-O benzyl group of 10a was explored. Surprisingly, hydrogenolysis of the benzyl ether using several palladium-carbon catalysts under a hydrogen atmosphere or in the presence of hydrogen transfer reagents (HCO₂H, HCO₂NH₄, cyclohexene etc.) was unsuccessful. In all cases, no reaction occurred and only the starting material was recovered. An attempt to cleave the benzyl ether using BCl₃ resulted in a complex mixture, affording the desired deprotected product 11 in only 10% yield. However, in contrast to these difficulties of cleaving the benzyl ether, removal of the MPM group from 10b was accomplished in high yield. After 6'deoxygenation of 6b was carried out by a sequence similar to that for 6a (70%), 10b was treated with DDQ in CH₂Cl₂-H₂O (9:1) to afford compound 11 in 93% yield.

The nucleoside 11 can be employed for the synthesis of plicacetin, one of the simplest structures in the amicetin family, according to the literature.³ Additionally, it is also possible to transform the 4-methoxy moiety of 11 into a keto or amino functionality by treatment with an acid or ammonia solution.^{5a}

In conclusion, we established a new synthetic route to 2',3',6'-trideoxy- β -D-erythro-hexopyranosyl pyrimidine nucleosides by employing the intramolecular glycosylation method, which afforded the deoxy- β -nucleosides in a completely stereoselective manner. In this reaction as well as at the following deprotection step, the MPM group was found to play an important role in obtaining the corresponding products in reasonable yields.

EXPERIMENTAL

All melting points are uncorrected. ¹H and ¹³C NMR spectra were obtained on a JEOL JNM-EX400 spectrometer in CDCl₃ with Me₄Si as an internal standard. *J* Values

SCHEME 3

are given in Hz. IR spectra were recorded on a Perkin-Elmer Model 1600 spectrophotometer. TLC was performed on plates coated with silica gel 60 F₂₅₄ (Merck). For column chromatography, Wakogel C-300 (Wako Chemicals) was used. All solvents used in the reactions were distilled from an appropriate drying agent and stored over molecular sieves.

Phenyl 4,6-di-*O* -a cetyl-2,3-dideoxy-1-thio-α,β-D-erythro-hexopyranoside (3). To a solution of methyl 4,6-di-*O*-acetyl-2,3-dideoxy-D-erythro-hexofuranoside⁶ (2) (6.55 g, 26.6 mmol) and PhSSiMe₃ (5.6 ml, 30 mmol) in CH₂Cl₂ (270 mL) under Ar was added BF₃·OEt₂ (3.6 mL, 30 mmol)⁷ at room temperature, and the reaction mixture was stirred for 17 h. The solution was poured into saturated aqueous NaHCO₃ and extracted with CH₂Cl₂. The organic layer was dried over MgSO₄ and then the solvent evaporated. The residue was chromatographed on silica gel (hexane-AcOEt, 4:1) to provide 3 (7.01 g, 81%) as a partially white solid. ¹H NMR δ 1.80–1.96 (m, 1H), 2.00–2.20 (m, 2H), 2.03 (s), 2.04 (s), 2.08 (s), 2.20–2.35 (m, 1H), 3.65–3.72 (m), 4.09 (dd, J=2.2, 12.0), 4.17–4.25 (m), 4.28 (dd, J=5.9, 12.7), 4.46–4.54 (m), 4.66–4.83 (m), 4.79 (dd, J=2.2, 11.5, C-1(β)), 5.60 (d, J=5.4, C-1(α)), 7.20–7.40 (m, 3H), 7.45–7.58 (m, 2H). IR (neat) 2955, 1742, 1654, 1440, 1372, 1237, 1043, 996 cm⁻¹. Anal. Calcd. for C₁₆H₂₀O₅S: C, 59.24; H, 6.21. Found: C, 59.49; H, 6.24.

Phenyl 4-O-benzyl-2,3-dideoxy-1-thio- α , β -D-erythro-hexopyranoside (4a). To a solution of 3 (1.19 g, 3.7 mmol) in MeOH (10 mL) was added NaOMe (23 mg), and the reaction mixture was stirred at room temperature for 1d. After neutralization with ion-exchange resin (Dowex 50W, H⁺-form), the mixture was filtered and evaporated to dryness. The residue was dissolved in dry pyridine (14 mL), and then TrCl (3.3 g, 11.8 mmol) was added to the solution. After 17 h, the reaction mixture was poured into ice-cold water. The solution was extracted with CH₂Cl₂, and the organic layer was washed successively with dilute HCl, water, and saturated aqueous NaHCO₃. The organic layer was dried over MgSO₄, evaporated, and the residue was then

chromatographed on silica gel (hexane-AcOEt, 4:1) to provide phenyl 6-*O*-trityl-2,3-dideoxy-1-thio- α , β -D-*erythro*-hexopyranoside (1.58 g, 83%) as a light yellow oil. ¹H NMR δ 1.50–1.90 (m), 1.95–2.20 (m), 3.25–3.73 (m), 4.22 (ddd, *J*=4.9, 5.4, 9.3), 4.75 (dd, *J*=2.4, 13.7, *C*-1(β)), 5.50 (d, *J*=4.4, C-1 (α)), 7.20–7.57 (m, 20H). IR (neat) 3448, 3058, 3007, 2930, 1490, 1448, 1070 cm⁻¹.

To a solution of the resulting 6-O-trityl derivative (1.58 g, 3.3 mmol) in DMF (33 mL) was added NaH (50% in oil, 0.32 g, 6.6 mmol) at 0 °C. After 1 h, BnBr (0.78 mL, 6.6 mmol) was added dropwise to the reaction mixture at 0 °C, and the mixture was stirred at room temperature for 16 h The reaction mixture was poured into saturated aqueous NH₄Cl, extracted with ether, and washed with water. The organic layer was dried over MgSO₄ and then evaporated to dryness. The residue was dissolved in BuOH (10 mL) and CF₃CO₂H (3.3 mL) was added to the solution. After 8 h, the solution was neutralized with saturated aqueous NaHCO3 and BuOH was removed by evaporation. The residue was extracted with CHCl₃. The organic layer was dried over MgSO₄ and then the solvent evaporated. The residue was chromatographed on silica gel (hexane-AcOEt, 4:1) to provide **4a** (955 mg, 88%) as an oil. ¹H NMR δ 1.50–1.90 (m), 2.00–2.20 (m), 2.30– 2.40 (m), 3.35–3.55 (m), 3.67–3.87 (m), 3.90 (dd, J=2.9, 11.7), 4.21 (ddd, J=3.9, 4.4, 9.3), 4.48 (d, J=11.2), 4.47 (d, J=11.7), 4.62 (d, J=11.2), 4.68 (d, J=11.2), 4.84 (dd, $J=2.0, 11.2, C-1(\beta)$), 5.54 (d, $J=3.9, C-1(\alpha)$), 7.20–7.50 (m, 10H). IR (neat) 3457, 2929, 2870, 1534, 1215, 1091 cm⁻¹. Anal. Calcd. for C₁₉H₂₂O₃S: C, 69.06; H, 6.71. Found: C, 69.34; H, 6.96.

Phenyl 4-*O*-*p*-methoxybenzyl-2,3-dideoxy-1-thio-α,β-D-*erythro*-hexopyranoside (4b). As described above, deacetylation of 3 (6.77 g, 20.9 mmol) afforded crude phenyl 2,3-dideoxy-1-thio-D-*erythro*-hexopyranoside, which was dissolved in DMF (200 mL) under Ar, and *p*-anisaldehyde dimethylacetal (3.9 mL, 23 mmol) and HBF₄-OEt₂ (3.2 mL, 22 mmol)¹¹ were then added at room temperature. After 1 h, Et₃N (3.1 mL) was added to the reaction mixture, and the solvent was removed by evaporation. The residue was dissolved in ether and the organic layer was washed with water and then dried over MgSO₄. After evaporation, the residue was chromatographed on silica gel (hexane-AcOEt, 9:1) to provide phenyl 4,6-di-*O*-*p*-methoxybenzylidene-2,3-dideoxy-1-thio-D-*erythro*-hexopiranoside (7.06 g, 94%) as a white solid. ¹H NMR δ 1.73–2.10 (m, 2H), 2.12–2.37 (m, 2H), 3.48 (ddd, J=4.9, 9.8, 9.8), 3.55–3.90 (m), 3.79 (s), 3.80 (s), 4.14 (dd, J=4.9, 10.3), 4.27 (dd, J=4.9, 10.8), 4.35 (ddd, J=4.9, 9.8, 9.8), 4.90 (dd, J=2.0, 11.7, C-1(β)), 5.51 (s), 5.54 (s), 5.56 (d, J=4.9, C-1(α)), 6.85–6.95 (m, 2H), 7.20–7.37 (m, 3H), 7.37–7.55 (m, 4H). IR (KBr) 2935, 2891, 1615, 1518, 1368, 1248, 1179, 1089, 1032 cm⁻¹.

To a solution of the resulting p-methoxybenzylidene derivative (2.15 g, 6.0 mmol) in MeCN (120 mL) was added NaBH₃CN (2.30 g, 36 mmol) and molecular sieves 3A (1.2 g) under Ar. After 30 min, the solution was cooled to 0 °C, and Me₃SiCl (4.6 mL, 36 mmol) in MeCN (30 mL) was then added.⁸ The reaction mixture was stirred at room temperature for 2 h, filtered through Celite and poured into ice-cold saturated aqueous NaHCO₃. The aqueous layer was extracted with CH₂Cl₂. The organic layer was washed with saturated aqueous NaHCO₃, dried over MgSO₄, and then evaporated. The residue was chromatographed on silica gel (hexane-AcOEt, 4:1) to provide **4b** (1.88 g, 87%) as a colorless oil, along with 0.13 g (6%) of 6-O-MPM derivative. ¹H NMR δ 1.50–1.90 (m), 2.05–2.20 (m), 2.27–2.37 (m), 3.36–3.51 (m), 3.68–3.93 (m), 3.80 (s), 3.81 (s), 4.18 (ddd, J=3.4, 4.9, 8.3), 4.41 (d, J=11.2), 4.44 (d, J=11.2), 4.55 (d, J=11.2), 4.61 (d, J=11.2), 4.83 (dd, J=2.0, 11.7, C-1(β)), 5.54 (d, J=3.4, C-1(α)), 6.86–6.97 (m, 2H), 7.20–7.38 (m, 5H), 7.40–7.55 (m, 2H). IR (neat) 3442, 1661, 1627, 1543, 1486, 1250, 1094 cm⁻¹. Anal. Calcd. for C₂₀H₂₄O₄S: C, 66.64; H, 6.71. Found: C, 66.81; H, 6.85.

Phenyl 4-O-benzyl-6-O-(4-methoxypyrimidin-2-yl)-2,3-dideoxy-1-thio-α,β-D-erythro-hexopyranoside (5a). To a solution of 4a (425 mg, 1.29 mmol) in DMF (6.4 mL) was added NaH (50% in oil, 124 mg, 2.6 mmol) at 0 °C under Ar. After 30 min, the solution was cooled to -50 °C and then 2-chloro-4-methoxypyrimidine⁹ in DMF (2 mL) was added. The reaction mixture was stirred at -20 °C for 15 h. The reaction was quenched with saturated aqueous NH4Cl. The solution was extracted with CHCl₃. The organic layer was dried over MgSO₄ and evaporated. The residue was chromatographed on silica gel (hexane-AcOEt, 5:1) to provide 5a (433 mg, 77%) as a colorless oil. ¹H NMR δ 1.55-1.94 (m), 2.05-2.28 (m), 2.33-2.42 (m), 3.50-3.63 (m), 3.68-3.85 (m), 3.93 (s), 3.95 (s), 4.40-4.73 (m), 4.82 (dd, J=2.0, 11.3, C-1(β)), 5.58 (d, J=3.9, C-1(α)), 6.35 (d, J=5.9, 1H), 7.15-7.38 (m, 8H), 7.40-7.56 (m, 2H), 8.16 (d, J=5.4), 8.18 (d, J=5.4). IR (neat) 2948, 1584, 1574, 1418, 1377, 1284, 1098, 1085 cm⁻¹.

Phenyl 4-O-p-methoxybenzyl-6-O-(4-methoxypyrimidin-2-yl)-2,3-dideoxy-1-thio-α,β-D-erythro-hexopyranoside (5b). According to the above procedure, starting with 4b (1.43 g, 4.0 mmol) in DMF (40 mL), NaH (60% in oil, 0.32 g, 8 mmol) and 2-chloro-4-methoxypyrimidine (1.16 g, 8 mmol), 5b (1.30 g, 71%) was obtained as a colorless oil after chromatography on silica gel (hexane-AcOEt, 4:1). 1 H NMR δ 1.52–1.65 (m), 1.74–1.90 (m), 2.40–2.39 (m), 3.48–3.59 (m), 3.66–3.75 (m), 3.77 (s), 3.78 (s), 3.93 (s), 3.95 (s), 4.38–4.70 (m), 4.81 (dd, J=2.0, 11.7, C-1(β)), 5.57 (d, J=4.4, C-1(α)), 6.35 (d, J=5.4), 6.37 (d, J=5.4), 6.79 (d, J=8.8), 6.82 (d, J=8.8), 7.14–7.31 (m, 5H), 7.44–7.55 (m, 2), 8.16 (d, J=5.4), 8.18 (d, J=5.4). IR (neat) 2946, 1584, 1574, 1514, 1418, 1284, 1248, 1085 cm⁻¹.

1-(4-O-Benzyl-2,3-dideoxy-β-D-erythro-hexopyranosyl)-4-methoxy-

pyrimidine (6a). To a solution of **5a** (158 mg, 0.36 mmol) in MeCN (90 mL) under Ar, powdered molecular sieves 4A (0.9 g) were added. After 15 h, the solution was cooled to -20 °C, and Me₂S(SMe)BF₄ (80 mg, 0.40 mmol) was then added. After 5 h, aqueous NaOH (1M, 30 mL) was added to the reaction mixture, and the mixture was warmed to 0 °C for over 2 h. After saturated aqueous NH₄Cl was added, the solution was stirred at room temperature for 30 min, then filtered through Celite and extracted with CHCl₃. The organic layer was dried over MgSO₄ and evaporated. The residue was chromatographed on silica gel (hexane-AcOEt, 1:5) to provide **6a** (69 mg, 56%) as a white solid, along with 23% of **7a**. mp 154.5–156°C; ¹H NMR δ 1.39–1.51 (m, 1H), 1.66–1.79 (m, 1H), 2.16–2.25 (m, 1H), 2.32–2.42 (m, 1H), 3.40–3.50 (m, 1H), 3.60–3.68 (m, 1H), 3.72–3.80 (m, 1H), 3.87–4.00 (m, 1H), 3.95 (s, 3H), 4.49 (d, *J*=11.7, 1H), 4.67 (d, *J*=11.7 1H), 5.85 (dd, *J*=1.5, 10.8, 1H, *C*-1), 5.91 (d, *J*=7.3, 1H), 7.25–7.40 (m, 5H), 7.66 (d, *J*=7.3, 1H); ¹³C NMR δ 27.6, 30.2, 54.4, 62.6, 71.0, 72.2, 81.2, 82.9, 95.8, 127.8, 128.0, 128.5, 137.7, 142.1, 155.2, 171.5; IR (KBr) 3454, 2929, 1654, 1635, 1550, 1317, 1095 cm⁻¹.

1-(4-*O*-*p*-Methoxybenzyl-2,3-dideoxy-β-D-*erythro*-hexopyranosyl)-4-methoxypyrimidine (6b). According to the above procedure, starting with 5b (419 mg, 0.89 mmol) in MeCN (224 mL), powdered MS 4A (4.1 g), Me₂S(SMe)BF₄ (193 mg, 0.98 mmol) and aq. NaOH (90 mL), 6b (266 mg, 79%) was obtained as a white solid after chromatography on silica gel (AcOEt), along with 12% of 7b. mp 133–134 °C; ¹H NMR δ 1.37–1.52 (m, 1H), 1.64–1.77 (m, 1H), 1.83–2.01 (br, 1H), 2.20 (m, J=3.4, 6.4, 12.7, 1H), 2.35 (m, J=3.9, 7.8, 13.2 1H), 3.42 (ddd, J=4.4, 9.27, 10.7, 1H), 3.62 (ddd, J=3.4, 4.9, 8.3, 1H), 3.75 (dd, J=4.9, 12.2, 1H), 3.81 (s, 3H), 3.88 (dd, J=2.9, 11.7, 1H), 3.95 (s, 3H), 4.43 (d, J=11.2, 1H), 4.60 (d, J=11.2, 1H), 5.84 (dd, J=2.4, 8.3, 1H, C-1), 5.91 (d, J=7.8, 1H), 6.89 (d, J=8.8, 1H), 7.25 (d, J=8.8, 1H), 7.65 (d, J=7.3, 1H): ¹³C NMR δ 27.7, 30.3, 54.5, 55.3, 62.7, 70.7, 72.0, 81.2, 83.0, 96.0, 114.0, 129.5, 129.8, 142.2, 155.3, 159.5, 171.6; IR (KBr) 3442, 1662, 1627, 1543, 1486, 1106, 1094 cm⁻¹.

1-(4-O-Benzyl-6-phenylthio-2,3,6-trideoxy-β-D-erythro-

hexopyranosyl)-4-methoxypyrimidine (8a). To a solution of 6a (166 mg, 0.48 mmol) in pyridine (2.5 mL), PhSSPh (312 mg, 1.4 mmol) and Bu₃P (0.30 mL, 1.2 mmol) were added. After 4 h, the reaction mixture was poured into water and extracted with CHCl₃. The organic layer was dried over MgSO₄ and evaporated. The residue was chromatographed on silica gel (hexane-AcOEt, 1:1) to provide 8a. ¹H NMR δ 1.25–1.40 (m, 1H), 1.61–1.75 (m, 1H), 2.22–2.3 (m, 1H), 2.3–2.43 (m, 1H), 3.20 (dd, J=5.4, 14.2, 1H), 3.49 (dd, J=2.4, 14.2, 1H), 3.58–3.67 (m, 1H), 3.80–3.88 (m, 1H), 3.93 (s,

3H), 4.42 (d, *J*=11.7, 1H), 4.65 (d, *J*=11.7, 1H), 5.70 (dd, *J*=2.0, 10.3, 1H, *C*-1), 5.73 (d, *J*=7.3, 1H), 7.16 (d, *J*=7.8, 1H), 7.13–7.45 (m, 10H).

1-(4-*O*-Benzyl-6-iodo-2,3,6-trideoxy-β-D-erythro-hexopyranosyl)-4-methoxypyrimidine (9a). To a solution of 6a (97 mg, 0.28 mmol) in pyridine (2 mL), I_2 (86 mg, 0.34 mmol) and Ph_3P (89 mg, 0.34 mmol) were added. After 5 h, the reaction mixture was poured into aqueous $Na_2S_2O_3$ and extracted with CHCl₃. The organic layer was dried over MgSO₄ and evaporated. The residue was chromatographed on silica gel (0.3% MeOH/CHCl₃) to provide 9a. mp 146–149 °C; ¹H NMR δ 1.40–1.54 (m, 1H), 1.70–1.85 (m, 1H), 2.25–2.43 (m, 2H), 3.18 (ddd, J=3.4, 3.4, 8.8, 1H), 3.40–3.5 (m, 1H), 3.49 (dd, J=2.9, 10.7, 1H), 3.62 (dd, J=3.4, 10.7, 1H), 3.96 (s, 3H), 4.53 (d, J=11.2, 1H), 4.69 (d, J=11.2, 1H), 5.88 (dd, J=2.0, 10.8, 1H, C-1), 5.95 (d, J=7.7, 1H), 7.28–7.47 (m, 5H), 7.74 (d, J=7.3, 1H); ¹³C NMR δ 8.7, 26.9, 30.3, 54.5, 71.1, 75.4, 77.9, 82.7, 96.1, 128.0, 128.6, 137.6, 142.4, 155.2, 171.1; IR (KBr) 1670, 1633, 1542, 1481, 1316, 1190, 1125, 1101 cm⁻¹.

1-(4-O-Benzyl-2,3,6-trideoxy-β-D-erythro-hexopyranosyl)-4-

methoxypyrimidine (10a). To a degassed solution of 9a (92 mg, 0.20 mmol) in toluene (4 mL), Bu₃SnH (65 μL, 0.24 mmol) and AIBN (1.7 mg, 0.01 mmol) were added under Ar, and the solution was heated under reflux. After 6 h, the solution was cooled to room temperature and the solvent evaporated. The residue was chromatographed on silica gel (hexane-AcOEt, 1:1) to provide 10a (44 mg, 67%) as a white solid. mp 102-104 °C; 1 H NMR δ 1.25–1.49 (m, 1H), 1.33 (d, J=5.9, 3H), 1.60–1.76 (m, 1H), 2.20–2.40 (m, 2H), 3.05–3.16 (m, 1H), 3.60–3.70 (m, 1H), 3.95 (s, 3H), 4.49 (d, J=11.7, 1H), 4.67 (d, J=11.7, 1H), 5.77 (dd, J=2.0, 10.7, 1H, C-1), 5.91 (d, J=7.3, 1H), 7.28–7.43 (m, 5H), 7.67 (d, J=7.3, 1H); 13 C NMR δ 18.7, 28.0, 30.9, 54.6, 71.3, 77.8, 77.9, 83.1, 95.9, 128.0×2, 128.7, 142.5, 155.5, 171.8; IR (KBr) 2931, 1670, 1636, 1546, 1479, 1304, 1095 cm⁻¹.

1-(4-O-p-Methoxybenzyl-6-iodo-2,3,6-trideoxy-β-D-erythro-

hexopyranosyl)-4-methoxypyrimidine (9b). According to the procedure described for 9a, starting with 6b (98 mg, 0.26 mmol), pyridine (2.6 mL), I_2 (132 mg, 0.52 mmol), and PPh₃ (136 mg, 0.52 mmol), 9b (107 mg, 85%) was obtained as a white solid after chromatography on silica gel (0.2% MeOH/CHCl₃). mp 107–108 °C; ¹H NMR δ 1.36–1.50 (m, 1H), 1.67–1.81 (m, 1H), 2.25–2.37 (m, 2H), 3.16 (ddd, J=2.9, 3.4, 8.8, 1H), 3.40 (ddd, J=4.4, 8.8, 10.8, 1H), 3.48 (dd, J=2.9, 10.7, 1H), 3.59 (dd, J=3.9, 10.7, 1H), 3.81 (s, 3H), 3.96 (s, 3H), 4.47 (d, J=10.7, 1H), 4.61 (d, J=10.7, 1H), 5.87 (dd, J=2.0, 10.3, 1H, S=2.1, 5.95 (d, S=7.3, 1H), 6.90 (d, S=8.3, 2H), 7.27 (d, S=8.4, 2H), 7.74 (d, S=7.3, 1H); S=7.70 NMR δ 8.7, 27.0, 30.4, 54.5, 55.3, 71.0, 75.1, 77.9, 82.7, 96.1, 114.0, 129.6, 129.7, 142.4, 155.2, 159.5, 171.7; IR (KBr) 1670, 1640, 1544, 1515, 1486, 1328, 1306, 1251, 1094, 1014 cm⁻¹.

1-(4-*O*-*p*-Methoxybenzyl-2,3,6-trideoxy-β-D-*erythro*-hexopyranosyl)-4-methoxypyrimidine (10b). According to the procedure described for 10a, starting with 9b (319 mg, 0.66 mmol), toluene (13 mL), Bu₃SnH (0.35 mL, 1.3 mmol), and AIBN (5 mg, 0.03 mmol), 10b (195mg, 82%) was obtained as a solid after chromatography on silica gel (hexane-AcOEt, 1:1). mp 96–97 °C; ¹H NMR δ 1.25–1.47 (m, 1H), 1.31 (d, J=5.9, 3H), 1.60–1.73 (m, 1H), 2.20–2.34 (m, 2H), 3.07 (ddd, J=4.4, 8.8, 10.7, 1H), 3.63 (dq, J=5.9, 8.8, 1H), 3.81 (s, 3H), 3.95 (s, 3H), 4.42 (d,

J=4.4, 8.8, 10.7, 1H), 3.63 (dq, J=5.9, 8.8, 1H), 3.81 (s, 3H), 3.95 (s, 3H), 4.42 (d, J=11.2, 1H), 4.59 (d, J=11.2, 1H), 5.75 (dd, J=2.0, 10.3, 1H, C-1), 5.90 (d, J=7.8, 1H), 6.89 (d, J=8.8, 2H), 7.26 (d, J=7.8, 2H), 7.67 (d, J=7.3, 1H); ¹³C NMR δ 18.4, 27.8, 30.7, 54.4, 55.3, 70.8, 77.6, 82.8, 95.7, 113.8, 129.4, 130.1, 142.3, 155.3, 159.3, 171.5; IR (KBr) 1673, 1634, 1542, 1485, 1252, 1086 cm⁻¹.

1-(2,3,6-Trideoxy-β-D-erythro-hexopyranosyl)-4-methoxypyrimidine

(11). To a solution of 10b (67 mg, 0.19 mmol) in CH₂Cl₂ (1.8 mL)-water (0.2 mL) was added DDQ (53 mg, 0.22 mmol) at room temperature. ¹² After 6 h, saturated aqueous NaHCO₃ was added to the reaction mixture. The solution was extracted with 10% *i*-PrOH/AcOEt. The organic layer was dried over Na₂SO₄ and then evaporated. The residue was chromatographed on silica gel (AcOEt) to provide 11 (41 mg, 93%) as a solid. mp 137–138 °C; ¹H NMR δ 1.34 (d, *J*=5.9, 3H), 1.43–1.54 (m, 1H), 1.67–1.79 (m, 1H), 2.16–2.24 (m, 2H), 3.28–3.37 (m, 1H), 3.52 (dq, *J*=6.4, 9.3, 1H), 3.96 (s, 3H), 5.77 (dd, *J*=2.0, 10.3, 1H, *C*-1), 5.92 (d, *J*=7.3, 1H), 7.69 (d, *J*=7.8, 1H); ¹³C NMR δ 18.1, 30.9, 31.3, 54.4, 71.0, 78.8, 82.9, 95.9, 142.3, 155.4, 171.5; IR (KBr) 3326, 2948, 1632 (br), 1546, 1480, 1422, 1348, 1310, 1223, 1107, 1088, 1055, 1034 cm⁻¹.

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REFERENCES

- 1. For a review, Lerner, L. M. Chem. Nucleosides Nucleotides 1991, 2, 27-79.
- 2. For recent studies, (a) Itoh, J.; Miyadoh, S. J. Antibiotics 1992, 45, 846–853 and references cited therein. (b) Shiomi, K.; Haneda, K.; Tomoda, H.; Iwai, Y.; Omura, S. J. Antibiotics 1994, 47, 782–786.
- Stevens, C. L.; Nielsen, N. A.; Blumbergs, F.; Taylor, K. G. J. Am. Chem. Soc. 1964, 86, 5695-5697.
- 4. Böhringer, M.; Roth, H.-J.; Hunziker, J.; Göbel, M.; Krishnan, R.; Giger, A.; Schweizer, B.; Schreiber, J.; Leumann, C.; Eschenmoser, A. Helv. Chim. Acta 1992, 75, 1416-1477.

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- (a) Sujino, K.; Sugimura, H. Chem. Lett. 1993, 1187-1190. (b) Sujino, K.; Sugimura, H. Tetrahedron Lett. 1994, 35, 1883-1886. (c) Sugimura, H.; Motegi, M.; Sujino, K. Nucleosides Nucleotides 1995, 14, 413-416.
- 6. Ferrier, R. J.; Prasad, N. J. Chem. Soc. (C) 1969, 570-575.
- Nicolaou, K. C.; Seitz, S. P.; Papahatjis, D. P. J. Am. Chem. Soc. 1983, 105, 2430-2434.
- 8. Johansson, R.; Samuelsson, B. J. Chem. Soc., Perkin Trans. I 1984, 2371-2374.
- 9. Katritzky, A. R.; Baykut, G.; Rachwal, S.; Szafran, M.; Caster, K. C.; Eyler, J. J. Chem. Soc., Perkin Trans. II 1989, 1499-1506.
- 10. Nakagawa, I.; Aki, K.; Hata, T. J. Chem. Soc. Perkin Trans. I 1983, 1315-1318.
- Albert, R.; Dax, K.; Pleschko, R.; Stütz, A. E. Carbohydr. Res. 1985, 137, 282–290.
- 12. Oikawa, Y.; Yoshioka, T.; Yonemitsu, O. Tetrahedron Lett. 1982, 23, 885-888.